



## King's Research Portal

DOI:

[10.1155/2020/4321419](https://doi.org/10.1155/2020/4321419)

*Document Version*

Publisher's PDF, also known as Version of record

[Link to publication record in King's Research Portal](#)

*Citation for published version (APA):*

Huang, X. F., Sun, L., Zhang, C., Zhou, Z., Chen, H., Zhang, L., Brown, M. A., & Xia, X. (2020). Whole-Exome Sequencing Reveals a Rare Missense Variant in SLC16A9 in a Pedigree with Early-Onset Gout. *BioMed Research International*, 2020, 1-6. [4321419]. <https://doi.org/10.1155/2020/4321419>

### Citing this paper

Please note that where the full-text provided on King's Research Portal is the Author Accepted Manuscript or Post-Print version this may differ from the final Published version. If citing, it is advised that you check and use the publisher's definitive version for pagination, volume/issue, and date of publication details. And where the final published version is provided on the Research Portal, if citing you are again advised to check the publisher's website for any subsequent corrections.

### General rights

Copyright and moral rights for the publications made accessible in the Research Portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognize and abide by the legal requirements associated with these rights.

- Users may download and print one copy of any publication from the Research Portal for the purpose of private study or research.
- You may not further distribute the material or use it for any profit-making activity or commercial gain
- You may freely distribute the URL identifying the publication in the Research Portal

### Take down policy

If you believe that this document breaches copyright please contact [librarypure@kcl.ac.uk](mailto:librarypure@kcl.ac.uk) providing details, and we will remove access to the work immediately and investigate your claim.

## Research Article

# Whole-Exome Sequencing Reveals a Rare Missense Variant in *SLC16A9* in a Pedigree with Early-Onset Gout

Xiu-Feng Huang,<sup>1</sup> Li Sun,<sup>2</sup> Chunwu Zhang,<sup>3</sup> Zhenni Zhou,<sup>4</sup> Hui Chen,<sup>5</sup> Linhua Zhang,<sup>6</sup> Matthew A. Brown,<sup>7,8</sup> and Xiaoru Xia<sup>ID</sup><sup>2</sup>

<sup>1</sup>Institute of Health and Biomedical Innovation, Queensland University of Technology, Translational Research Institute, Brisbane, QLD, Australia

<sup>2</sup>Department of Rheumatology, First Affiliated Hospital of Wenzhou Medical University, Wenzhou, China

<sup>3</sup>Department of Injury Orthopaedics, First Affiliated Hospital of Wenzhou Medical University, Wenzhou, China

<sup>4</sup>Department of Internal Medicine, Yueqing People's Hospital, Yueqing, China

<sup>5</sup>Department of Nephrology, Wenzhou Central Hospital, Wenzhou, China

<sup>6</sup>Department of Clinical Laboratory, Yuhuan People's Hospital, Yuhuan, China

<sup>7</sup>Centre for Precision Medicine, First Affiliated Hospital of Wenzhou Medical University, Wenzhou, China

<sup>8</sup>Guy's & St Thomas' NHS Foundation Trust and King's College London NIHR Biomedical Research Centre, London, UK

Correspondence should be addressed to Xiaoru Xia; [xrx7799@163.com](mailto:xrx7799@163.com)

Received 17 June 2019; Revised 28 December 2019; Accepted 10 January 2020; Published 31 January 2020

Academic Editor: Rachid Tazi-Ahnini

Copyright © 2020 Xiu-Feng Huang et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Gout is a common inflammatory arthritis triggered by monosodium urate deposition after longstanding hyperuricemia. In the general community, the disease is largely polygenic in genetic architecture, with many polymorphisms having been identified in gout or urate-associated traits. In a small proportion of cases, rare high penetrant mutations associated with monogenic segregation of the disease in families have been demonstrated to be disease causative. In this study, we recruited a two-generation pedigree with early-onset gout. To elucidate the genetic predisposition, whole-exome sequencing (WES) was performed. After comprehensive variant analyses and cosegregation testing, we identified a missense variant (c.277C>A, p.L93M) in *SLC16A9*, an extremely rare variant in genetic databases. Moreover, *in silico* assessments showed strong pathogenicity. This variant cosegregated with the disease phenotype perfectly in the family and is located in a highly conserved functional domain. A few studies supported our results of the association between *SLC16A9* and gout and serum urate levels. In conclusion, we provide the first evidence for the association of rare missense in *SLC16A9* with early-onset gout. These findings not only expand our current understanding of gout but also may have further implications for the treatment and prevention of gout.

## 1. Introduction

Gout is a common inflammatory arthritis caused by the deposition of monosodium urate (MSU) crystals in and around the joints following longstanding hyperuricemia [1]. It affects 1–2% of adults in developed countries [1–3] and has a prevalence of 1.14% in eastern China [4]. Similar to other complex phenotypes, gout results from the interplay between inherited genetic risk variants and environmental factors [5]. Genome-wide association studies (GWAS) have

confirmed the importance of genetic basis in gout. Several genetic loci have been associated with gout, such as *ABCG2*, *ALDH16A1*, *BCAS3*, *RFX3*, *KCNQ1*, *ATXN2*, *CUX2*, *GCKR*, *PDZK1*, *CNTN5*, and mitochondrial genetic variation [6–14]. For example, *ABCG2* dysfunctional variants have a strong impact on the progression of hyperuricemia. The most common dysfunction variant rs2231142 (p.Q141K) increases the risk of gout and hyperuricemia, significantly influences the age of onset of gout, and is highly associated with a familial gout history [15]. Moreover, *ABCG2*

dysfunction was reported as a strong independent risk factor for pediatric-onset hyperuricemia/gout [16].

Notably, almost all these loci identified in gout GWAS were also associated with serum urate levels, indicating the shared genetic basis between gout and serum urate concentrations [5]. This is mainly because elevated serum urate levels are a critical risk factor for gout onset [17]. However, as GWAS for gout have been relatively limited in size and power compared with the GWAS of serum urate levels, less is known about the specific genetic contribution to gout as opposed to genetic associations of hyperuricemia. Association with gout at most urate-associated loci is still unclear.

*SLC16A9* encodes monocarboxylate transporter 9 (MCT9), a member of solute carrier (SLC) superfamily that comprises more than 400 transporters [18]. *SLC16A9* is ubiquitously expressed, including at particularly high levels in the kidney [19, 20]. Recent GWAS and meta-analysis revealed a significant association between polymorphisms of *SLC16A9* and serum urate concentrations [7, 21, 22]. However, a role for variants of *SLC16A9* and gout itself has not been demonstrated [7, 11]. To date, the transport substrate of MCT9 is still unknown and the function of *SLC16A9* remains poorly understood, especially its potential association with gout.

Despite dozens of genetic loci identified in gout or urate-associated traits, little is known about the genetic aetiology of patients presenting with early-onset gout (EOG), which was defined as before the age of 40 years [23–26]. Previous studies have reported rs2231142 (Q141K) in *ABCG2* as a genetic factor in early-onset gout [16, 27, 28]. In this study, we investigated a two-generation pedigree with early-onset gout. To elucidate the genetic predisposition, whole-exome sequencing (WES) was performed, and we identified a rare missense mutation (c.277C>A, p.L93M) in *SLC16A9*, providing new evidence for the association of *SLC16A9* with gout.

## 2. Materials and Methods

**2.1. Participant Recruitment.** This study conformed to the tenets of the Declaration of Helsinki and was approved by the Ethics Committee of the First Affiliated Hospital of Wenzhou Medical University. Informed consent was obtained from the patient. Patients were clinically evaluated by rheumatologist according to the 2015 gout classification criteria by an American College of Rheumatology/European League against Rheumatism Collaborative Initiative [29, 30]. Peripheral blood samples were collected from patients and unaffected family members, from which genomic DNA was extracted.

**2.2. Whole-Exome Sequencing.** Whole-exome sequencing (WES) was performed on the proband. Briefly, genomic DNA was sheared into 200- to 250-base pair (bp) fragments using a Covaris S220 ultrasonicator. Then the fragments were ligated with adapters to both ends, amplified by ligation-mediated polymerase chain reaction, purified, and hybridized. Nonhybridized fragments were washed out.

Enrichment of the DNA libraries was performed using the Exome Enrichment V5 Kit (Agilent Technologies, Palo Alto, CA, USA) according to the manufacturers' protocol. Subsequently, enriched DNA libraries were sequenced on a HiSeq X Ten sequencer (Illumina, San Diego, CA, USA). All raw sequencing data were processed according to a customized bioinformatics pipeline described previously [31]. After the quality control test, the reads were mapped to the reference human genome (hg19) using SOAPaligner software and further visualized using the SplicingViewer software [32]. SNV and Indel calls as well as annotation were performed using GATK tool and mirTrios with integrated ANNOVAR tool [33].

**2.3. Variant Analyses and Identification.** We used the following databases for selecting rare variants as an initial filtration: Genome Aggregation Database (<http://gnomad.broadinstitute.org/>), Exome Aggregation Consortium (ExAC, <http://exac.broadinstitute.org/>), NHLBI Exome Sequencing Project (ESP, <http://evs.gs.washington.edu/EVS/>), and 1000 Genome (<http://www.1000genomes.org>). Variants with a minor allele frequency of over 0.01 in any of these databases were discarded. The effects of the candidate variants were assessed using in silico prediction programs. Missense variants were analyzed by M-CAP (<http://bejerano.stanford.edu/mcap/>), Polyphen-2 (<http://genetics.bwh.harvard.edu/pph2/>), and MutationTaster (<http://mutationtaster.org/>). Direct Sanger sequencing was then used to confirm the segregated variants in the present family, using an ABI 3500 Genetic Analyzer (Applied Biosystems, Carlsbad, CA, USA).

**2.4. *SLC16A9* Amplification and Genotyping.** An additional cohort of unrelated cases ( $n = 30$ ) with gout was recruited, and their DNA was submitted for Sanger sequencing. Primers were designed to amplify all coding regions and the intron-exon boundaries of the *SLC16A9* gene. The PCR products were purified and sequenced on an ABI 3500 Genetic Analyzer (Applied Biosystems, Carlsbad, CA, USA).

## 3. Results

**3.1. Clinical Observations.** The proband was a 25-year-old male from a Han Chinese family, having suffered his first gout flare at the age of 20. His father was also diagnosed with gout having the first gout flare at the age of 25, whereas all the other family members were unaffected (Table 1 and Figure 1(a)).

Both proband and his affected father experienced recurrent acute monoarticular arthritis affecting the first metatarsophalangeal joint (MTP1) and/or knee starting at 20 and 25 years of age, respectively. The symptoms generally started at night and peaked within 24 hours, preventing walking and could not bear touch. The symptomatic course lasted no more than one week. The symptoms typically completely resolved within one or two days after taking nonsteroidal anti-inflammatory drug and colchicine. Patients have normal intelligence and are competent of the job.

TABLE 1: Summary of clinical observations of the participants in this study.

ID	Gender	Age (y)	SUA	HUA	SCr	BUN	FEUa	UPH	Onset age (y)	Arthritis	Tophi	TG	TC	Obesity	HBP	HG	Obesity
II:1	F	63	257	–	51	6.8		6.5	–	–	–	–	–	–	+	–	–
II:4	M	48	581	+	69	4.2	3.77	6.0	25	+	–	–	+	+	+	+	+
II:5	F	46	305	–	44	6.0		5.5	–	–	–	+	–	+	–	–	+
III:1	M	45	321	–	50	5.3		6.0	–	–	–	–	–	–	–	–	–
III:2	M	25	517	+	71	5.1	4.63	5.0	20	+	–	–	–	–	–	–	–

SUA, serum uric acid,  $\mu\text{mol/l}$ ; HUA, hyperuricemia; SCr, serum creatine,  $\mu\text{mol/l}$ ; BUN, blood urea nitrogen,  $\text{mmol/l}$ ; FEUa, fractional excretion of uric acid,%; (hyperuricemia: male  $> 420 \mu\text{mol/l}$ ; female  $> 360 \mu\text{mol/l}$ ); UPH, urine PH; TG, triglyceride; TC, total cholesterol; HBP, high blood pressure; HG, hyperglycemia.

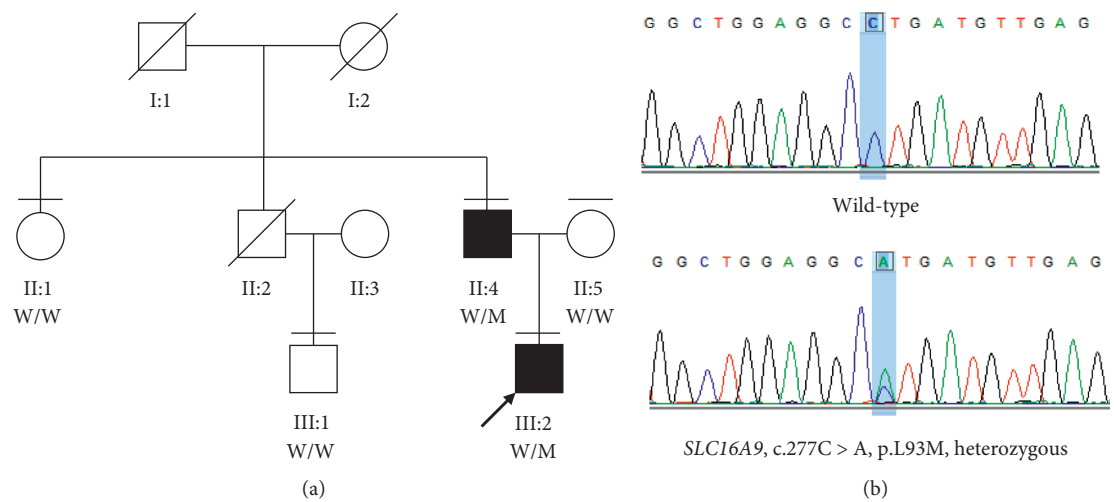


FIGURE 1: Identification of SLC16A9 missense in the family with early-onset gout. (a) Pedigree and cosegregation results. Affected individual is represented as a filled square. Normal individuals are shown as empty symbols. (b) Sanger sequencing confirmed the segregation of the rare missense variant, c.277C>A (p.L93M).

The muscle tension and renal function are normal, and no urate nephrolithiasis has been found. In both cases, serum uric acid was increased but not achieved to an extremely high level (see in Table 1), and the fractional excretion of uric acid (FEUa) was decreased (the normal range for FEUa is 7%–12%) [30, 34], consistent with renal underexcretion (RUE) gout. So the purine overproduction gout (HGPR1 deficiency, PRPS1 superactivity) was excluded in the family. The detailed clinical information is summarized in Table 1.

**3.2. Genetic Assessments.** To reveal the genetic predisposition, WES was performed on the proband (III:2). The mean read depth for the WES was  $>100\times$  and the coverage of the targeted regions ( $>1\times$ ) reached  $>99\%$ . Variant analyses and a step-by-step filtering strategy by combination of minor allele frequency, in silico assessments, gene function, and cosegregation analysis were carried out [35–37]. A total of six candidate variants were submitted for cosegregation analysis and only one survived, a rare missense mutation (c.277C>A, p.L93M) in SLC16A9. This is an extremely rare variant (rs550527563) in all of the databases (Table 2). For example, the allele frequency is 0.0032% (8 in 251050) and 0.0033% (4 in 120986) in gnomAD and ExAC, respectively, while it is absent in ESP (Table 2). All these alleles are from Asian, while it is absent in Caucasians. Moreover, in silico

assessments showed strong pathogenicity for this variant including the M-CAP, a newly developed tool for variants with uncertain significance in clinical exomes at high sensitivity [38]. Importantly, segregation testing in all available family members indicated that L93M cosegregated with the disease phenotype in this pedigree (Figures 1(a) and 1(b)). Both patients harbor a heterozygous variant while the healthy individuals do not have the nucleotide change. The variant c.277C>A results in a switch from leucine to methionine in the major facilitator superfamily (MFS) domain (Figure 2(a)). Multiple orthologous sequence alignment revealed that leucine at position 93 is in a highly conserved region across different species (Figure 2(b)).

The probability of being loss-of-function (LoF) intolerant (pLI) is 0.64, and the expected number of LoF is 11.7 while the observed number is only 2, suggesting the probability of being a functionally important variant [39]. Expanded screening of SLC16A9 in a cohort of 30 patients with gout failed to identify any additional rare variants in this gene. Taken together, WES revealed a putative causal variant in SLC16A9 in a family with early-onset gout.

#### 4. Discussion

In the present study, we recruited an unusual pedigree with early-onset gout. It is reasonable to speculate that it is

TABLE 2: Variant identified in patients with early-onset gout.

ID	Variant	Type	Frequency (allele count)				In silico assessments			
			gnomAD	ExAC	ESP	1K	Polyphen-2	MutationTaster	LRT	M-CAP
II:4	c.277C>A, p.L93M	Hetero	0.0032% (8)	0.0033% (4)	0	0.04% (2)	Damaging	Damaging	Damaging	Possibly pathogenic
III:2	c.277C>A, p.L93M	Hetero	0.0032% (8)	0.0033% (4)	0	0.04% (2)	Damaging	Damaging	Damaging	Possibly pathogenic

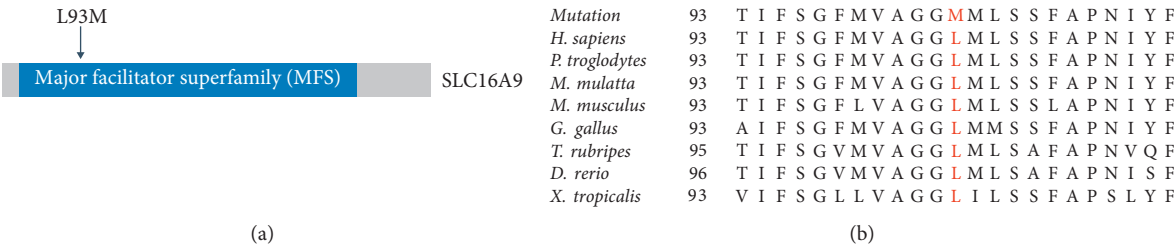


FIGURE 2: (a) Domain structure of SLC16A9 and location of L93M variant. (b) Conservation analyses of the mutated residues 93 in *SLC16A9* across different species.

possibly caused by rare monogenic variants because of two reasons, the family history and early-onset age. Firstly, this gout pedigree exhibited an autosomal dominant-like trait, consistent with a monogenic aetiology. More importantly, both proband and his affected father suffered first gout flare at early age. Epidemiological studies show that gout incidence increases with age until the age of 70 years and that onset before the age of 40 years is unusual [40, 41]. A few studies have demonstrated that complex disease with early-onset age could be caused by monogenic inheritance of mutated genes [42–44]. Therefore, aiming to investigate potential causal gene in this pedigree, we used WES which has proven to be highly robust and efficient in the identification of disease-causing genes in monogenic conditions or complex disorders [45]. Using this approach, we identified a rare missense in *SLC16A9* gene by the comprehensive analyses including allele frequency, in silico assessments, gene function, and cosegregation analysis.

A few studies supported our results of the association between *SLC16A9* and gout. The first evidence reported by Kolz et al. observed a SNP in *SLC16A9*, rs12356193, was significantly associated with serum uric acid levels by a meta-analysis of 28,141 individuals of European descent ( $P = 1.1 \times 10^{-8}$ ) [21]. Then the locus was successfully replicated in a cohort of 7,795 individuals [22]. Nakayama et al. investigated the relationship between another common variant (rs2242206) and gout. They found that the  $P$  value was significant in renal overload gout (ROL), but not with all gout susceptibility [46]. Subsequently, Köttgen et al. confirmed the *SLC16A9* locus was associated with serum urate concentrations (rs1171614,  $P = 2.3 \times 10^{-28}$ ), but showed only nominal association with gout (rs1171614,  $P = 1.7 \times 10^{-2}$ ) [7]. Phipps-Green et al. tested 28 loci for association with gout in 1536 cases with gout and 2645 controls. At *SLC16A9*, the observed association with gout was restricted to the lower Polynesian ancestry group (rs12356193,  $P = 0.006$ ) [11]. Of note, the relationship between GWAS signals and genes underlying Mendelian

phenotypes has been observed [47, 48]. Thus, it is reasonable to find rare pathogenic variants in GWAS signals. In addition to these genetic association studies, several studies also provided functional evidence. *SLC16A9* is ubiquitously expressed and is especially expressed at a high level in the kidney [19, 20]. *ALDH16A1* gene is associated with serum uric acid levels and gout, and RNA sequencing in the kidney of wild-type (WT) and *Aldh16a1* knockout (KO) mice revealed changes in *Slc16a9* are localized to the apical membrane of the proximal convoluted tubule cells and influence uric acid homeostasis [49]. These findings suggested the potential role of *SLC16A9* in the aetiology of gout.

However, there are two main limitations in this study. Firstly, no functional genomics studies were performed in the present study. Experimental validations are essential to determine if interesting variants are indeed responsible for clinical symptoms [50, 51]. For example, a recent study demonstrates the rare variants of *ABCG2* at both the clinical level and the functional level by complex approach [52]. Second is the lack of independent replication family. The genetic screening of *SLC16A9* in gout pedigrees is required in the future studies. The copy number variations (CNVs) are not considered in this study [53].

In conclusion, we provide the first evidence for the association of rare missense in *SLC16A9* with early-onset gout. These findings not only expand our current understanding of gout, but also may have further implications for the treatment and prevention of gout.

### Data Availability

Summary data are available from the corresponding author on request.

### Conflicts of Interest

The authors declare no conflicts of interest.



## Authors' Contributions

X.X. and X.F.H. conceived and designed the experiments; X.X. and L.S. recruited patients and collected samples; X.F.H., L.S., and C.Z. performed the experiments, analyzed data, and contributed equally to this work; Z.Z., H.C., and L.Z. contributed reagents/materials/analysis tools; X.X., X.F.H., and M.A.B. wrote and revised the manuscript. M.A.B. and X.X. also contributed equally to this work. X.F.H., L.S., and C.Z. contributed equally to this work.

## Acknowledgments

This study was supported by the Zhejiang Provincial Natural Science Foundation (LY20H100001), National Natural Science Foundation of China (31771390), and Wenzhou Science and Technology Bureau (Y20180129).

## References

- [1] P. Richette and T. Bardin, "Gout," *The Lancet*, vol. 375, no. 9711, pp. 318–328, 2010.
- [2] K. L. Wallace, A. A. Riedel, N. Joseph-Ridge, and R. Wortmann, "Increasing prevalence of gout and hyperuricemia over 10 years among older adults in a managed care population," *The Journal of Rheumatology*, vol. 31, no. 8, pp. 1582–1587, 2004.
- [3] L. Annemans, E. Spaepen, M. Gaskin et al., "Gout in the UK and Germany: prevalence, comorbidities and management in general practice 2000-2005," *Annals of the Rheumatic Diseases*, vol. 67, no. 7, pp. 960–966, 2008.
- [4] Z. Miao, C. Li, Y. Chen et al., "Dietary and lifestyle changes associated with high prevalence of hyperuricemia and gout in the Shandong coastal cities of Eastern China," *The Journal of Rheumatology*, vol. 35, no. 9, pp. 1859–1864, 2008.
- [5] T. J. Major, N. Dalbeth, E. A. Stahl, and T. R. Merriman, "An update on the genetics of hyperuricaemia and gout," *Nature Reviews Rheumatology*, vol. 14, no. 6, pp. 341–353, 2018.
- [6] P. Sulem, D. F. Gudbjartsson, G. B. Walters et al., "Identification of low-frequency variants associated with gout and serum uric acid levels," *Nature Genetics*, vol. 43, no. 11, pp. 1127–1130, 2011.
- [7] A. Köttgen, E. Albrecht, A. Teumer et al., "Genome-wide association analyses identify 18 new loci associated with serum urate concentrations," *Nature Genetics*, vol. 45, no. 2, pp. 145–154, 2013.
- [8] C. Li, Z. Li, S. Liu et al., "Genome-wide association analysis identifies three new risk loci for gout arthritis in Han Chinese," *Nature Communications*, vol. 6, no. 1, p. 7041, 2015.
- [9] H. Matsuo, K. Yamamoto, H. Nakaoka et al., "Genome-wide association study of clinically defined gout identifies multiple risk loci and its association with clinical subtypes," *Annals of the Rheumatic Diseases*, vol. 75, no. 4, pp. 652–659, 2016.
- [10] A. Nakayama, H. Nakaoka, K. Yamamoto et al., "GWAS of clinically defined gout and subtypes identifies multiple susceptibility loci that include urate transporter genes," *Annals of the Rheumatic Diseases*, vol. 76, no. 5, pp. 869–877, 2017.
- [11] A. J. Phipps-Green, M. E. Merriman, R. Topless et al., "Twenty-eight loci that influence serum urate levels: analysis of association with gout," *Annals of the Rheumatic Diseases*, vol. 75, no. 1, pp. 124–130, 2016.
- [12] Y. Kawamura, H. Nakaoka, A. Nakayama et al., "Genome-wide association study revealed novel loci which aggravate asymptomatic hyperuricaemia into gout," *Annals of the Rheumatic Diseases*, vol. 78, no. 10, pp. 1430–1437, 2019.
- [13] A. L. Gosling, J. Boockock, N. Dalbeth et al., "Mitochondrial genetic variation and gout in Maori and Pacific people living in Aotearoa New Zealand," *Annals of the Rheumatic Diseases*, vol. 77, no. 4, pp. 571–578, 2018.
- [14] A. Tin, J. Marten, V. L. Halperin Kuhns et al., "Target genes, variants, tissues and transcriptional pathways influencing human serum urate levels," *Nature Genetics*, vol. 51, no. 10, pp. 1459–1474, 2019.
- [15] B. Stiburkova, K. Pavelcova, J. Zavada et al., "Functional non-synonymous variants of ABCG2 and gout risk," *Rheumatology*, vol. 56, no. 11, pp. 1982–1992, 2017.
- [16] B. Stiburkova, K. Pavelcova, M. Pavlikova, P. Ješina, and K. Pavelka, "The impact of dysfunctional variants of ABCG2 on hyperuricemia and gout in pediatric-onset patients," *Arthritis Research & Therapy*, vol. 21, no. 1, p. 77, 2019.
- [17] P. L. Riches, A. F. Wright, and S. H. Ralston, "Recent insights into the pathogenesis of hyperuricaemia and gout," *Human Molecular Genetics*, vol. 18, no. R2, pp. R177–R184, 2009.
- [18] A. César-Razquin, B. Snijder, T. Frappier-Brinton et al., "A call for systematic research on solute carriers," *Cell*, vol. 162, no. 3, pp. 478–487, 2015.
- [19] A. P. Halestrap and D. Meredith, "The SLC16 gene family—from monocarboxylate transporters (MCTs) to aromatic amino acid transporters and beyond," *Pflügers Archiv European Journal of Physiology*, vol. 447, no. 5, pp. 619–628, 2004.
- [20] A. P. Halestrap, "The SLC16 gene family—structure, role and regulation in health and disease," *Molecular Aspects of Medicine*, vol. 34, no. 2-3, pp. 337–349, 2013.
- [21] M. Kolz, T. Johnson, S. Sanna et al., "Meta-analysis of 28,141 individuals identifies common variants within five new loci that influence uric acid concentrations," *PLoS Genetics*, vol. 5, no. 6, Article ID e1000504, 2009.
- [22] P. van der Harst, S. J. L. Bakker, R. A. de Boer et al., "Replication of the five novel loci for uric acid concentrations and potential mediating mechanisms," *Human Molecular Genetics*, vol. 19, no. 2, pp. 387–395, 2010.
- [23] P. Richette, M. Doherty, E. Pascual et al., "2016 updated EULAR evidence-based recommendations for the management of gout," *Annals of the Rheumatic Diseases*, vol. 76, no. 1, pp. 29–42, 2017.
- [24] K.-H. Yu and S. F. Luo, "Younger age of onset of gout in Taiwan," *Rheumatology*, vol. 42, no. 1, pp. 166–170, 2003.
- [25] B. Zhang, W. Fang, X. Zeng et al., "Clinical characteristics of early- and late-onset gout: a cross-sectional observational study from a Chinese gout clinic," *Medicine (Baltimore)*, vol. 95, no. 47, p. e5425, 2016.
- [26] T. Pascart, L. Norberciak, H. K. Ea, P. Guggenbuhl, and F. Lioté, "GOSPEL 4—patients with early onset gout develop earlier severe joint involvement and metabolic comorbid conditions," *Arthritis Care and Research (Hoboken)*, vol. 71, no. 7, pp. 986–992, 2019.
- [27] M. Cleophas, L. Joosten, L. Stamp, N. Dalbeth, O. Woodward, and T. Merriman, "ABCG2 polymorphisms in gout: insights into disease susceptibility and treatment approaches," *Pharmacogenomics and Personalized Medicine*, vol. 10, pp. 129–142, 2017.
- [28] H. Matsuo, H. Tomiyama, W. Satake et al., "ABCG2 variant has opposing effects on onset ages of Parkinson's disease and gout," *Annals of Clinical and Translational Neurology*, vol. 2, no. 3, pp. 302–306, 2015.
- [29] T. Neogi, T. L. T. A. Jansen, N. Dalbeth et al., "2015 gout classification criteria: an American College of Rheumatology/

- European League against rheumatism collaborative initiative," *Arthritis & Rheumatology*, vol. 67, no. 10, pp. 2557–2568, 2015.
- [30] T. Neogi, T. L. T. A. Jansen, N. Dalbeth et al., "2015 gout classification criteria: an American College of Rheumatology/European League against rheumatism collaborative initiative," *Annals of the Rheumatic Diseases*, vol. 74, no. 10, pp. 1789–1798, 2015.
- [31] T. Wang, Q. Liu, X. Li et al., "RRBS-analyser: a comprehensive web server for reduced representation bisulfite sequencing data analysis," *Human Mutation*, vol. 34, no. 12, pp. 1606–1610, 2013.
- [32] Q. Liu, C. Chen, E. Shen, F. Zhao, Z. Sun, and J. Wu, "Detection, annotation and visualization of alternative splicing from RNA-Seq data with SplicingViewer," *Genomics*, vol. 99, no. 3, pp. 178–182, 2012.
- [33] J. Li, Y. Jiang, T. Wang et al., "mirTrios: an integrated pipeline for detection of de novo and rare inherited mutations from trios-based next-generation sequencing," *Journal of Medical Genetics*, vol. 52, no. 4, pp. 275–281, 2015.
- [34] Q. H. Li, J. J. Liang, L. X. Chen et al., "Clinical characteristics and renal uric acid excretion in early-onset gout patients," *Zhonghua Nei Ke Za Zhi*, vol. 57, no. 3, pp. 185–190, 2018, in Chinese.
- [35] X. F. Huang, Z. Q. Huang, D. Lin et al., "Unraveling the genetic cause of a consanguineous family with unilateral coloboma and retinoschisis: expanding the phenotypic variability of RAX mutations," *Scientific Reports*, vol. 7, no. 1, p. 9064, 2017.
- [36] X.-F. Huang, F. Huang, K.-C. Wu et al., "Genotype-phenotype correlation and mutation spectrum in a large cohort of patients with inherited retinal dystrophy revealed by next-generation sequencing," *Genetics in Medicine*, vol. 17, no. 4, pp. 271–278, 2015.
- [37] X.-F. Huang, J. Wu, J.-N. Lv, X. Zhang, and Z.-B. Jin, "Identification of false-negative mutations missed by next-generation sequencing in retinitis pigmentosa patients: a complementary approach to clinical genetic diagnostic testing," *Genetics in Medicine*, vol. 17, no. 4, pp. 307–311, 2015.
- [38] K. A. Jagadeesh, A. M. Wenger, M. J. Berger et al., "M-CAP eliminates a majority of variants of uncertain significance in clinical exomes at high sensitivity," *Nature Genetics*, vol. 48, no. 12, pp. 1581–1586, 2016.
- [39] M. Lek, K. J. Karczewski, E. V. Minikel et al., "Analysis of protein-coding genetic variation in 60,706 humans," *Nature*, vol. 536, no. 7616, pp. 285–291, 2016.
- [40] M. Doherty, "New insights into the epidemiology of gout," *Rheumatology*, vol. 48, no. Suppl 2, pp. ii2–ii8, 2009.
- [41] C.-F. Kuo, M. J. Grainger, W. Zhang, and M. Doherty, "Global epidemiology of gout: prevalence, incidence and risk factors," *Nature Reviews Rheumatology*, vol. 11, no. 11, pp. 649–662, 2015.
- [42] M. Barbier, D. Wallon, and I. Le Ber, "Monogenic inheritance in early-onset dementia: illustration in Alzheimer's disease and frontotemporal lobar dementia," *Gériatrie et Psychologie Neuropsychiatrie du Vieillessement*, vol. 16, no. 3, pp. 289–297, 2018.
- [43] Z.-B. Jin, J. Wu, X.-F. Huang et al., "Trio-based exome sequencing arrests de novo mutations in early-onset high myopia," *Proceedings of the National Academy of Sciences*, vol. 114, no. 16, pp. 4219–4224, 2017.
- [44] V. Bansal, J. Gassenhuber, T. Phillips et al., "Spectrum of mutations in monogenic diabetes genes identified from high-throughput DNA sequencing of 6888 individuals," *BMC Medicine*, vol. 15, no. 1, p. 213, 2017.
- [45] J. Wu, E. Shen, D. Shi, Z. Sun, and T. Cai, "Identification of a novel Cys146X mutation of SOD1 in familial amyotrophic lateral sclerosis by whole-exome sequencing," *Genetics in Medicine*, vol. 14, no. 9, pp. 823–826, 2012.
- [46] A. Nakayama, H. Matsuo, T. Shimizu et al., "Common missense variant of monocarboxylate transporter 9 (MCT9/SLC16A9) gene is associated with renal overload gout, but not with all gout susceptibility," *Human Cell*, vol. 26, no. 4, pp. 133–136, 2013.
- [47] J. X. Chong, K. J. Buckingham, S. N. Jhangiani et al., "The genetic basis of mendelian phenotypes: discoveries, challenges, and opportunities," *The American Journal of Human Genetics*, vol. 97, no. 2, pp. 199–215, 2015.
- [48] M. K. Freund, K. S. Burch, H. Shi et al., "Phenotype-specific enrichment of mendelian disorder genes near GWAS regions across 62 complex traits," *The American Journal of Human Genetics*, vol. 103, no. 4, pp. 535–552, 2018.
- [49] G. Charkoftaki, Y. Chen, M. Han et al., "Transcriptomic analysis and plasma metabolomics in Aldh16a1-null mice reveals a potential role of ALDH16A1 in renal function," *Chemico-Biological Interactions*, vol. 276, pp. 15–22, 2017.
- [50] X. F. Huang, L. Xiang, X. L. Fang et al., "Functional characterization of CEP250 variant identified in nonsyndromic retinitis pigmentosa," *Human Mutation*, vol. 40, no. 8, pp. 1039–1045, 2019.
- [51] X. F. Huang, L. Xiang, W. Cheng et al., "Mutation of IPO13 causes recessive ocular coloboma, microphthalmia, and cataract," *Experimental & Molecular Medicine*, vol. 50, no. 4, p. 53, 2018.
- [52] Y. Toyoda, A. Mančíková, V. Krylov et al., "Functional characterization of clinically-relevant rare variants in ABCG2 identified in a gout and hyperuricemia cohort," *Cells*, vol. 8, no. 4, p. 363, 2019.
- [53] X.-F. Huang, J.-Y. Mao, Z.-Q. Huang et al., "Genome-wide detection of copy number variations in unsolved inherited retinal disease," *Investigative Ophthalmology & Visual Science*, vol. 58, no. 1, pp. 424–429, 2017.